Two New Alkaloids from Incarvillea mairei var. grandiflora

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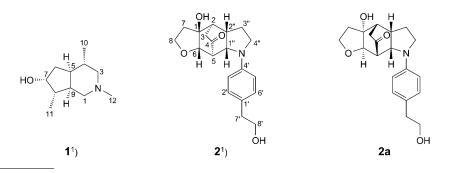
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Two new alkaloids, isoincarvilline (1) and incargranine A (2), together with two known ones, were isolated from the 80% EtOH extract of the whole plant of *Incarvillea mairei* var. *grandiflora*. Their structures were identified on the basis of their spectroscopic analysis.

Introduction. – Incarvillea mairei var. grandiflora (WEHRHAHN) GRIERSON (Bignoniaceae), a beautiful mountain flower, is mainly distributed in the mountains of Yunnan, Sichuan, and Qinghai provinces [1][2]. The investigations on Incarvillea species have let to the isolation of many novel actinidine-type monoterpenoid alkaloids with strong antinociceptive activity [3–14]. Although no evidences were found for the use of the title plant in traditional Chinese medicine, the structural diverse and novel alkaloids from Invarvillea species still attracted our interest and decided us to perform the pharmacological and phytochemical investigations of I. mairei var. grandiflora. In our investigation of the components from this plant, two new alkaloids, isoincarvilline¹) (1) and incargranine A¹) (2), together with two known ones, were isolated from the 80% EtOH extract of the whole plant of I. mairei var. grandiflora. Their structures were elucidated on the basis of their spectroscopic analysis.



1) Arbitrary atom numbering; for systematic names, see Exper. Part.

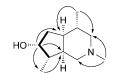
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Results and Discussion. – The CHCl₃ fraction of the 80% EtOH extract of *I. mairei* var. grandiflora was purified by repeated column chromatography to afford four compounds. On the basis of physical and spectroscopic analysis, including 2D-NMR techniques (HMQC, HMBC, and NOESY), the structures of two new compounds were determined and named isoincarvilline¹) (1) and incargranine A¹) (2), and two known alkaloids were deduced to be β -skytanthine (=(4*S*,4a*R*,7*S*,7a*S*)-octahydro-2,4,7-trimethyl-1*H*-cyclopenta[*c*]pyridine) [15] and incarvine C (=(2*E*)-3-(4-hydroxy-3-methoxyphenyl)prop-2-enoic acid (4*S*,4a*R*,6*S*,7*R*,7a*S*)-octahydro-2,4,7-trimethyl-1*H*-cyclopenta[*c*]pyridin-6-yl ester) [10] by comparing their spectroscopic data with those reported in the literature.

Compound **1** was isolated as colorless needles which showed a positive reaction to the *Dragendorff* reagent, indicating that **1** was an alkaloid. The molecular formula of **1** was determined to be $C_{11}H_{21}NO$ by the quasimolecular-ion peak $[M+H]^+$ at m/z 184.1700 in the HR-ESI-MS (positive mode). Comparison of its ¹H- and ¹³C-NMR data (*Table*) with those of the known β -skytanthine [15] and incarvilline (=(4R,4aS,6R, 7S,7aR)-octahydro-2,4,7-trimethyl-1*H*-cyclopenta[*c*]pyridin-6-ol) [9] suggested that **1** has the same planar structure as the latter, consistent with the HMBC and COSY (*Fig. 1*). An X-ray diffraction analysis of **1** (*Fig. 2*) was performed to establish the relative α -configurations of H–C(5), H–C(9), Me(10), Me(11), and OH–C(7). Consequently, the structure of isoincarvilline (**1**) was determined.

Table. ¹*H*- and ¹³*C*-*NMR* Data of Compounds 1^1) and 2^1). δ in ppm, J in Hz.

1			2					
	$\delta(\mathrm{H})^{\mathrm{a}})$	$\delta(C)^b)$		$\delta(H)^{c})$	$\delta(C)^d)$		$\delta(H)^{c})$	$\delta(C)^d)$
$H_a - C(1)$	2.77 $(d, J = 12.0)$	55.3 (t)	C(1)		81.7 (s)	H-C(5')	6.62 (d, J = 8.4)	115.5 (d)
$H_{\beta}-C(1)$	2.11 (dd , $J = 3.6, 12.0$)		H-C(2)	2.37–2.39 (<i>m</i>)	42.7 (<i>d</i>)	H-C(6')	7.05 $(d, J = 8.4)$	130.6 (<i>d</i>)
$H_a - C(3)$	2.68 $(d, J = 10.2)$	63.2 (<i>t</i>)	$H_a - C(3)$	2.49 (dd , J = 3.3, 18.9)	36.7 (<i>t</i>)	CH ₂ (7')	2.70 $(t, J = 7.2)$	39.4 (<i>t</i>)
$H_{\beta}-C(3)$	1.41 – 1.47 (overlap)		$H_b-C(3)$	2.31 (dd , $J = 3.1, 18.9$)		CH ₂ (8')	3.67(t, J = 7.2)	64.6 (<i>t</i>)
H-C(4)	1.31–1.34 <i>(m)</i>	34.3 (<i>d</i>)	C(4)		214.1 (s)	H-C(1")	4.06 (dd, J = 3.0, 8.6)	60.1 (<i>d</i>)
H-C(5)	1.41 – 1.47 (overlap)	42.0 (<i>d</i>)	H-C(5)	3.21 (dd, J = 3.6, 3.0)	53.9 (d)	H-C(2")	3.34-3.36 (<i>m</i>)	36.1 (<i>d</i>)
$H_a - C(6)$	1.68 $(dd, J = 8.4, 14.4)$	39.3 (<i>t</i>)	H-C(6)	3.83 (d, J = 3.6)	86.7 (<i>d</i>)	$H_a - C(3'')$	2.15–2.21 (<i>m</i>)	27.4 (<i>t</i>)
$H_{\beta}-C(6)$	1.86 (dd , $J = 6.0, 14.4$)		CH ₂ (7)	1.97–2.00 (<i>m</i>)	39.6 (<i>t</i>)	$H_b - C(3'')$	1.92–1.95 (<i>m</i>)	
H-C(7)	4.12-4.14 (<i>m</i>)	74.6 (d)	$H_a - C(8)$	3.89-3.95 (<i>m</i>)	68.2 (t)	$H_a - C(4'')$	3.40-3.48 (<i>m</i>)	51.2 (t)
H-C(8)	1.99 - 2.05 (m)	38.7 (d)	$H_b - C(8)$	3.78-3.83 (<i>m</i>)		$H_{b}-C(4'')$	3.13-3.18 (<i>m</i>)	
H-C(9)	1.73 $(t, J = 5.4)$	43.5 (d)	C(1')		129.1 (s)			
Me(10)	0.76 (d, J = 6.6)	17.7(q)	H-C(2')	7.05 $(d, J = 8.4)$	130.6 (d)			
Me(11)	0.93 (d, J = 6.0)	11.8(q)	H-C(3')	6.61 (d, J = 8.4)	115.5 (d)			
MeN	2.18 (s)	46.7 (q)	C(4′)		146.8 (s)			
^a) At 600 MHz in CDCl ₃ . ^b) At 150 MHz in CDCl ₃ . ^c) At 600 MHz in CD ₃ OD. ^d) At 150 MHz in CD ₃ OD.								



¹H,¹H-COSY \rightarrow HMBC Fig. 1. Key ¹H,¹H-COSY and HMBC of compound **1**

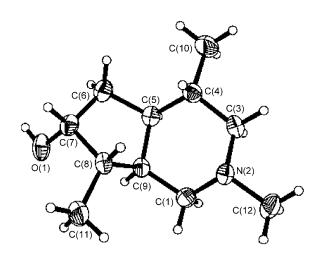


Fig. 2. *X-Ray crystallographic structure of* 1¹). The crystallographic data of 1 have been deposited with the *Cambridge Crystallographic Data Centre* as supplementary publication number CCDC-691814. Copies of the data can be obtained, free of charge, *via* http://www.ccdc.cam.ac.uk/data_request/cif.

The ¹H-NMR spectrum of **1** showed two Me *d* at $\delta(H) 0.93$ (J = 6.0 Hz) and 0.76 (J = 6.6 Hz), indicating the presences of two MeCH moieties. A *s* at $\delta(H) 2.18$ was ascribed to an MeN group. The ¹³C-NMR spectrum exhibited resonances for 11 C-atoms (one MeN, two Me, three CH₂ (sp³), five CH (sp³)). These data suggested that **1** was the derivative of a monoterpenoid alkaloid, and shared the same skeleton as β -skytanthine [15]. In the HMBC plot, the H-atoms of MeN were correlated to C(1), and C(3) (*Fig.* 1). The Me groups at $\delta(H) 0.76$ (Me(10) and 0.93 (Me(11)) showed long-range correlations with C(3), C(4), and (5), and with C(7), C(8), and C(9), respectively. An oxygenated H-atom resonance at $\delta(H) 4.12 - 4.14$ exhibited COSY cross-peaks with the signals of H–C(6) and H–C(8), implying that an OH group may be attached at C(7) (*Fig.* 2). This was further confirmed by the HMBC of H–C(7) at $\delta(H) 4.12 - 4.14$ with C(5), C(9), and C(11). Compound **1** was then determined to have the same planar structure as the known compound incarvilline [9]. Although the NOESY correlations H–C(5)/H–C(9), Me(11)/H–C(9), and H–C(9)/Me(10) were observed, it was still difficult to determine the relative configuration. The X-ray diffraction analysis confirmed the suggested relative configuration (*Fig.* 2).

Compound **2**, a light yellow oil, showed a positive reaction to the *Dragendorff* reagent. The ESI-MS (positive mode) gave a quasimolecular-ion peak $[M + Na]^+$ at m/z 366.3. Combined with the ¹H- and ¹³C-NMR data (*Table*), its molecular formula was deduced to be C₂₀H₂₅NO₄ with 9 degrees of unsaturation. The HR-ESI-MS (negative mode; m/z 378.1469 ($[M + Cl]^-$)) further confirmed the above deduction. The NMR data of **2** were quite close to those of the previously reported biotransformation

product **2a** [16], implying that their structures may be very similar. The ¹H,H-COSY (*Fig. 3*), HMBC (*Fig. 3*) and NOESY data (*Fig. 4*) of **2** confirmed that incargranine A (**2**) is a stereoisomer of **2a**.

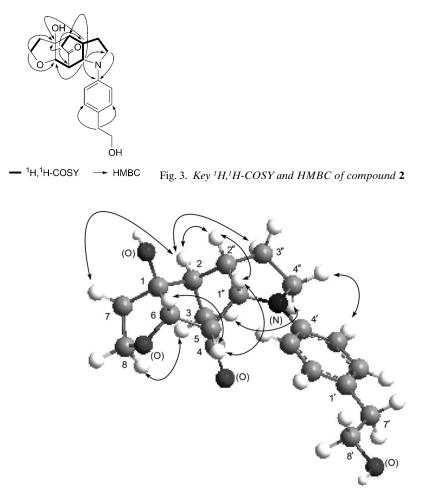


Fig. 4. Key NOESY correlations of Compound 2

The ¹H-NMR of **2** exhibited two *d* at δ (H) 7.05 (J = 8.4 Hz, 2 H) and 6.61 (J = 8.4 Hz, 2 H), suggesting the presence of a 4-substituted phenyl group. Two mutually coupling CH₂ at δ (H) 2.70 (t, J = 7.2 Hz) and 3.67 (t, J = 7.2 Hz) indicated a hydroxyethyl moiety. The HMBC of the CH₂ H-atoms at δ (H) 2.70 with C(1'), C(2'), and C(6') of the aryl group were consistent with a 4-(hydroxyethyl)phenyl group. The ¹³C-NMR displayed resonances for twenty C-atoms, including a C=O at δ (C) 214.1. Beside the aryl and C=O group, the remaining 4 degrees of unsaturation indicated a four-ring structural moiety. The ¹H,¹H-COSY plot showed the following correlations: CH₂(8)/CH₂(7), CH₂(3)/H-C(2)/H-C(2'')/H-C(1'')/H-C(5)/H-C(6), and H-C(2'')/CH₂(3'')/CH₂(4'') (*Fig.* 3). Moreover, the HMBCs CH₂(8)/C(1), and C(6), CH₂(7)/C(2) and C(6), H-C(6)/C(2), C(4), C(7), C(8), and C(1''), H-C(1'')/C(2),

C(4), C(6), C(3"), C(4'), and C(4"), H–C(2")/C(1), C(3), C(5), and C(4"), and CH₂(4")/C(1") and C(4'), indicated that **2** has the same planar structure as **2a**. In the NOESY plot of **2**, we observed the correlations H–C(2)/H–C(2") and CH₂(3"), and H–C(5)/H–C(1") (*Fig.* 4), indicating that the ethano bridge C(3)–C(4) was α -orientated and the bridgehead H–C(2) and H–C(5) β -orientated. Additionally, the relative β -configurations of H–C(6) and OH–C(1) were deduced from the NOESY correlations H–C(6)/H–C(5) and H–C(5) and CH₂(7)/CH₂(3).

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Experimental Part

General. Column chromatography (CC): silica gel (SiO₂, 200–300 mesh), *H60* (Qingdao Marine Chemical Plant, Qingdao, P. R. China); Sephadex LH-20 (Pharmacia Fine Chemicals, Piscataway, NJ, USA). TLC: pre-coated SiO₂ GF_{254} plates (Qingdao Marine Chemical Plant, Qingdao, P. R. China). Optical rotation: Perkin-Elmer-341 digital polarimeter (Perkin-Elmer, Norwalk, CT, USA); at 589 nm. IR: Bruker-Vector-22 spectrophotometer; KBr pellets; in cm⁻¹. ¹H-, ¹³C-, and 2D-NMR Spectra: Bruker-DRX-600 spectrometer; in CDCl₃; δ in ppm rel. to Me₄Si, J in Hz. MS: Agilent-1100-LC/MSD-Trap (ESI-MS) and Agilent Micro-Q-Tof (HR-ESI-MS) spectrometer; in m/z.

Plant Material. The whole plants of *I. mairei* var. *grandiflora* were collected in Zhongdian County, Yunnan Province, in late October 2006, and authenticated by Prof. *Li-Shan Xie* of the Kunming Institute of Botany, the Chinese Academy of Sciences. A voucher specimen (No. 2006101020) is deposited with the School of Pharmacy, Second Military Medical University.

Extraction and Isolation. The dried whole plants of *I. mairei* var. *grandiflora* (32.5 kg) were extracted 3 times with 80% EtOH under reflux. The EtOH extract was concentrated and the residue dissolved in 2% HCl soln. and filtered. The filtrate was adjusted to pH 9–10 by adding NH₄OH and then extracted with CHCl₃. The CHCl₃ fraction (120 g) was subjected to CC (SiO₂, petroleum ether/AcOEt (100:1 \rightarrow 5:1): *Fractions 1–5. Fr. 3* was purified by repeated CC (SiO₂; *Sephadex LH-20*, CHCl₃/MeOH 1:1): **1** (8 mg) and β -skytanthine (9 mg). *Fr. 5* (CHCl₃/MeOH 1:2) was purified by a similar procedure: **2** (7 mg) and incarvine C (8 mg).

Isoincarvilline (= rel-(4R,4aR,6S,7R,7aS)-*Octahydro-2,4,7-trimethyl-1*H-*cyclopenta[c]pyridin-6-ol*; 1): Colorless needles. $[a]_{2D}^{2D} = -77.3$ (c = 0.2335, CHCl₃). IR (KBr): 3215, 2962, 1460, 1376. ¹H- and ¹³C-NMR: *Table*. ESI-MS: 184 ($[M + H]^+$). HR-ESI-MS: 184.1700 ($[M + H]^+$, C₁₁H₂₂NO⁺; calc. 184.1701).

Incargranine A (=rel-(3aR,4S,4aR,7aR,8R,8aR)-Decahydro-3a-hydroxy-7-[4-(2-hydroxyethyl)-phenyl]-4,8-ethano-2H-furo[3,2-f]indol-9-one; **2**): Light yellow oil. $[a]_D^{2D} = +2$ (c = 0.1750, CHCl₃). ¹H- and ¹³C-NMR: Table. ESI-MS: 366.3 ($[M + Na]^+$), 378.2 ($[M + Cl]^-$). HR-ESI-MS: 378.1469 ($[M + Cl]^-$, $C_{20}H_{25}CINO_4^-$; calc. 378.1472).

REFERENCES

- [1] W. C. Wang, 'Flora of China', Science Press, Beijing, 1990, Vol. 69, p. 46.
- [2] K. Xu, Y. L. Liu, H. Hu, J. Yunnan Agric. Univ. 2005, 20, 559.
- [3] M. Nakamura, Y. M. Chi, J. Kinjo, W. M. Yan, T. Nohara, Phytochemistry 1999, 51, 595.
- [4] Y. M. Chi, F. Hashimoto, W. M. Yan, T. Nohara, *Phytochemistry* 1997, 46, 763.
- [5] Y. M. Chi, M. Nakamura, X. Y. Zhao, Nat. Prod. Res. Dev. (in Chinese) 2005, 17, 362.
- [6] T. F. Ji, X. Z. Fen, Chin. Tradit. Herb. Drugs 2002, 33, 967.
- [7] M. Nakamura, K. Kido, J. Kinjo, T. Nohara, Phytochemistry 2000, 53, 253.

- [8] Y. M. Chi, W. M. Yan, J. S. Li, Phytochemistry 1990, 29, 2376.
- [9] Y. M. Chi, W. M. Yan, D. C. Chen, N. Hiroshi, I. Yoichi, S. Ushio, Phytochemistry 1992, 31, 2930.
- [10] Y. M. Chi, F. Hashimoto, W. M. Yan, T. Nohara, Phytochemistry 1995, 39, 1485.
- [11] Y. M. Chi, F. Hashimoto, W. M. Yan, T. Nohara, *Phytochemistry* 1995, 40, 353.
- [12] Y. M. Chi, F. Hashimoto, W. M. Yan, T. Nohara, M. Yamashita, N. Marubayashi, *Chem. Pharm. Bull.* 1997, 45, 495.
- [13] M. Nakamura, Y. M. Chi, W. M. Yan, Y. Nakasugi, T. Yoshizawa, N. Irino, F. Hashimoto, J. Kinjo, T. Nohara, S. Sakurada, J. Nat. Prod. 1999, 62, 1293.
- [14] M. Nakamura, Y. M. Chi, W. M. Yan, A. Yonezawa, Y. Nakasugi, T. Yoshizawa, F. Hashimoto, J. Kinjo, T. Nohara, S. Sakurada, *Planta Med.* 2001, 67, 114.
- [15] A. P. Lins, J. D. Felicio, Phytochemistry 1993, 34, 876.
- [16] T. Hase, K. Ohtani, R. Kasai, Phytochemistry 1996, 41, 317.

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